

# Breakthroughs in the genetics of angle-closure glaucoma

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Angle closure glaucoma (ACG) is not widely known to be a familial condition, yet the recent explosion of genetic data and large scale genome wide investigations have confirmed at least 13 genetic loci associated with ACG [1], and provided some insight into the clinical and pathobiological mechanisms of the disease.

Primary angle closure glaucoma (PACG) is defined as iridotrabecular contact with evidence of elevated intraocular pressure (IOP), or peripheral anterior synechiae (PAS), and glaucomatous optic neuropathy (GON), with a reproducible visual field defect [2]. PACG is often diagnosed at the extremes of phenotypes when there is advanced chronic visual loss, or acute angle closure (AAC) [3]. Understanding the genetic aetiology for PACG can help identify at-risk individuals at earlier stages of the disease.

## Family history reporting

There are more people worldwide bilaterally blind from PACG than progressive open angle glaucoma (POAG) [4] but reported family history of PACG is less well studied. A clinic report from Brazil found 25% of their cases to have a reported family history of PACG [5]. A population survey in Harbin, Northeast China, found a positive family history to be a significant risk factor of PACG,

with an odds ratio of 65 under univariate logistic regression, or odds ratio of 1.65 in the multivariate model [6]. Amerasinghe et al. found that the relative risk of narrow angles to first degree family members is seven times higher than the general population of Singaporean Chinese [7]. Kong et al. compared 332 PAC patients, 228 POAG and 193 controls in Shanghai, China to investigate a glaucoma family history, finding that a parent with PACG contributed most to the family history of glaucoma in the PAC, compared to POAG where siblings and children contributed more to the family history of glaucoma [8]. In India, Kavitha et al. found that there was a 13.6 times greater odds of angle-closure in siblings of PAC/G subjects compared to open angle subjects in a hospital setting [9]. A greater age effect was observed for glaucoma in PACG.

## Accurate phenotyping is crucial

When Lowe et al. first described a series of families with angle-closure in Australia [10], little was known about the natural history of the disease. He noted that the rate of AAC increased as anterior chamber depth (ACDs) got shallower, but the co-inheritance of PACG in multiple family members was much less than angle-closure observed without glaucoma. He proposed a major environmental component to the disease

mechanism, while emphasising that familial PACG was rare but definitely observed [11]. Subsequent groups, such as Spaeth et al., Tomlinson et al. and Sihota et al. all reported small ocular biometry to be prominent in familial disease [12-14]. However, all these reports had low rates of AAC.

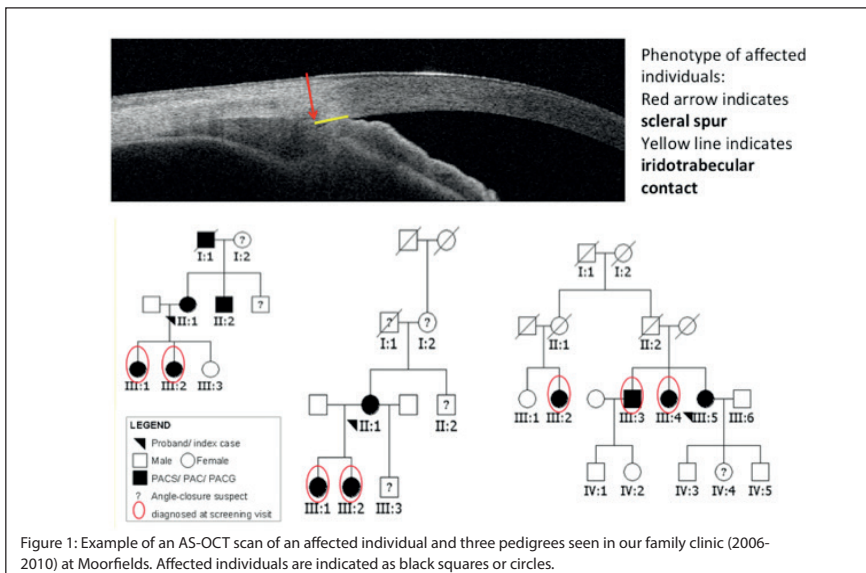
Using the Spaeth gonioscopic grading system, iris convexity was found to be a common feature amongst affected family members [13], and more recently anterior segment imaging with ultrasound biomicroscopy (UBM) and anterior segment optical coherence tomography (AS-OCT) have allowed detailed, objective quantification of angle closure traits such as thicker irides, anteriorly rotated ciliary bodies, and iris vault to be studied. Figure 1 shows a high resolution AS-OCT scan of an affected individual with iridotrabecular contact. Three pedigrees with multiple affected individuals are illustrated.

Sihota et al. found that shallower ACD, thicker lenses and shorter axial length (AL) segregated in family members affected with PACG compared to suspected and unaffected subjects [12]. Using the UBM, Etter et al. demonstrated a 50% rate of plateau iris configuration, in a small collection of predominantly white American families [15].

## Previous heritability and segregation analyses

The most recent segregation analysis reporting 114 PACG families from Chongqing, China [16], found a significant difference between gender groups. They concluded that the inheritance of shallow angles may be a sex-influenced trait with a reported female:male ratio of 2.87:1, consistent with the ratio found in other East Asian population studies of PACG [4,17]. Families with an unaffected (U) parent and an affected parent (A) accorded with an autosomal dominant hereditary trait, with the highest heritability found for female relatives with female probands [16].

Several investigators have provided data on the segregation of ocular biometry and PACG. Tomlinson and Leighton [14] examined 16 index PACG patients, their relatives (seven siblings, 14 offspring) and 49 controls, finding that the unaffected relatives and patients with PACG also had smaller corneal diameters, shallower (ACDs), thicker crystalline lenses and shorter axial



lengths compared to age-matched controls. However, those affected with PACG had more anteriorly positioned lenses, which were not observed in unaffected siblings and offspring. This finding was also observed by Rosengren [18] and Philips and Storey [19]. Törnquist from Sweden [20] found that eyes with PACG had ACDs of 1.0mm less than normal eyes, or two-thirds of the normal depth at corresponding ages.

Twin studies are often the starting point of dissecting genetic conditions, but there are no reported twin studies in PACG. A curious case report of two elderly identical twins who were in a heated argument and fist fight and both went into AAC together demonstrates that stress and other physiological triggers can influence disease presentation [21]. Hypermetropic refractive error is associated with angle-closure, but its contribution to PACG is likely to be incorporated with the heritability of ACD and AL [22-24].

### Genetic linkage and candidate gene studies

Once a clear phenotype is identified, such as AAC or PACG, genetic studies can be performed. Aung et al. reported genetic linkage in two large Chinese families, examining the sharing of phenotypic and genetic factors, more statistically found together in affected individuals than unaffected ones [25]. To date, this method of genetic linkage has not identified any causative genes for PACG. However, in more extreme forms of the disease, two genes, membrane frizzled related protein (*MFRP*, associated with retinitis pigmentosa) [26,27] and protease, serine 56 (*PRSS56*) [28] have been identified to be causative for autosomal recessive microphthalmia / nanophthalmia. Both genes were identified using linkage analysis.

Aung et al. also examined 108 patients with PACG (49 with AAC and 59 chronic cases) for *MFRP* and *CHX10* mutations [29]. *CHX10* is a homeobox-containing transcription factor critical for progenitor cell proliferation and bipolar cell determination in the developing retina. One potential disease causing variant, G243D was observed in *CHX10* in a patient with acute PACG who also had a *MFRP* R257H mutation. The clinical characteristics of the patient involved: an 82-year-old patient diagnosed at 75y with bilateral cup to disc ratio (CDR) of 0.8 and 360° of PAS and biometry of 21.05mm for AL and 2.09mm for ACD. This patient also carried an R46X polymorphism in the myocilin gene (*MYOC*) [30]. *MYOC* is an autosomal dominant genetic cause of POAG. The G243D mutation in *CHX10* may be pathogenic as it was not found in 400 normal controls, but it was not possible to show segregation of the G243D mutation in the subject's family members [29]. This was an example where sequence changes in three / multiple genes may each be contributing to

the disease in an individual patient with late onset symptomatic PACG.

### Genome wide association studies

PACG is clearly a complex disease with multifactorial inheritance, so it is no surprise that more success has been achieved with genome-wide association studies (GWAS). Using this methodology in 2012, three single-nucleotide polymorphisms (SNPs), rs11024102 in *PLEKHA7*, rs3753841 in *COL11A1* (a connective tissue gene) and rs1015213 between *PCMTD1* and *ST18* were found to be associated with AAC/PACG [3]. Day et al. compared these three SNPs for association to ACD, AL and corneal keratometry using an additive genetic model for each allele in the EPIC-eyes cohort. The presence of one A allele for rs1015213 was associated with a 0.07mm shallowing of ACD compared to wild type homozygotes after adjusting for the effects of age and sex [31]. The other two SNPs were not associated with the biometric characteristics tested. Little is known about the function of *PCMTD1* or *ST18*, the latter encodes for suppression of tumorigenicity 18 which is thought to modulate inflammation and apoptosis in fibroblasts [32] and has been reported as a breast cancer tumour suppressor gene [33].

The only GWAS study of ACD to date was reported by Vithana et al. where three well-characterised population based studies: Singapore Malay Eye Study (SiMES), Singapore Indian Eye Study (SINDI) and Beijing Eye Study (BES) with a total of 5308 participants were analysed. Two strongly associated intragenic SNPs on chromosome 3q27.1 were identified [34]. The gene of interest, *ABCC5* (rs1401999) was widely expressed and regulates cGMP levels in its role as an organic anion pump. They were able to replicate the finding in a further cohort of Chinese subjects but not in Caucasians. This SNP was only marginally associated with PACG in a group of patients. Interestingly, this gene was not found to be replicated in the meta-analysis paper presented by Rong et al. in 2016 [1]. Nongpiur et al. hypothesise that the *ABCC* family of genes may be involved in glaucoma as *ABCC4* has previously been found in the trabecular meshwork of a rabbit model with pigmentary glaucoma [34]. Perhaps overlapping anterior segment phenotypes will begin to unravel as more glaucoma genetic studies take place.

The most recent GWAS publication in PACG reported five further SNPs in *EPDR1*, *CHAT*, *GLIS3*, *FERMT2*, *DPM2-FAM10A* genes [35], and confirmed the original three loci (*PLEKHA7*, *COL11A1* and *PCTMD1-ST18*) in the 2012 Vithana paper [3] to be associated with AAC and PACG.

### Discussion

The strongest association signal to date is *PLEKHA7*. This cell-cell signaling gene lies close to the *NNO1* locus linked to extreme nanophthalmos where axial lengths of less

than 19mm and refractive errors of more than +7DS were observed [36]. The GWAS could not demonstrate an effect of *PLEKHA7* on axial biometry or nanophthalmos. However, it does suggest that physiological mechanisms that maintain homeostasis in epithelial and endothelial tissues can alter the susceptibility to AAC and PACG, i.e. more advanced stages of the disease.

In GWAS studies, often the only SNPs that are reported are ones that have reached genome-wide statistical significance set at a threshold of  $P < 5 \times 10^{-8}$ . In the 2012 Vithana paper [36], the fourth most significant locus was thioredoxin reductase 2 (*TXNRD2*), a gene involved in oxidative stress. Tissue expression studies and the relationship between this gene and its potential contribution to AAC or glaucoma have not yet been studied. The enzyme thioredoxin reductase (TR) is a dimeric NADPH-dependent FAD containing enzyme that catalyses the reduction of the active site of disulphide of thioredoxin and other substrates. It is a member of a family of pyridine nucleotide-disulphide oxidoreductases and a key enzyme in the regulation of the intracellular redox environment both in the cytosol and mitochondria [3]. This gene partially overlaps with the *COMT* gene on chromosome 22. The *COMT* gene encodes for catechol-O-methyltransferase (*COMT*) which catalyses the transfer of a methyl group from S-adenosylmethionine to catecholamines, including the neurotransmitters dopamine, epinephrine and norepinephrine. Oestrogens such as 17 $\beta$ -oestradiol, catabolised in hydroxylation reactions, are inactivated by methylation, a process catalysed by *COMT* [38]. This pathway is important in oestradiol metabolism. There is increasing evidence that the inheritance of particular variants in these oestrogen-metabolising genes can modulate the risk of hormone dependent disorders such as prostate, ovarian, lung and breast cancer [38]. Given the female preponderance to ACG, this area of work to understand the molecular genetic and biochemical mechanisms underlying PACG could lead to breakthroughs in our scientific understanding and delivery of novel therapies.

### Conclusion

There is emerging research in the genetics of ACG and keen phenotyping (skilled clinicians) in partnership with molecular biologists and statisticians can bridge the gaps in our knowledge of the disease. Figure 2 simplifies the phenotypes that have been discussed in this article. Even without genetic data, ophthalmic practitioners can recommend screening of family members, and proactively teach and learn gonioscopy, as accurate elucidation of clinical signs and appropriate management is essential for the diagnosis and prognosis of patients with AAC and PACG.

## PACG / AAC Candidate Genes

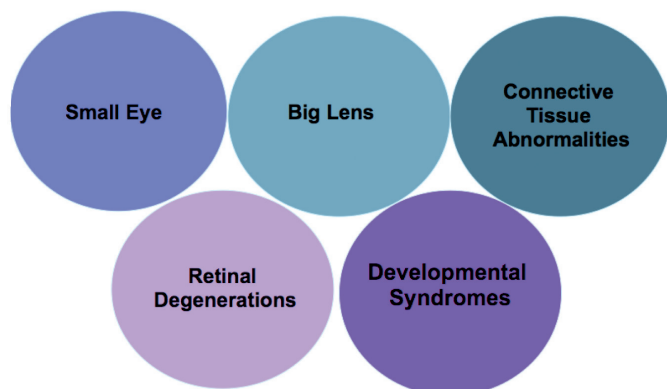


Figure 2: Phenotypes to look out for when considering a genetic cause for angle closure glaucoma.

### References

- Rong SS, Tang FY, Chu WK, et al. Genetic associations of primary angle-closure disease: a systematic review and meta-analysis. *Ophthalmology* 2016;**123**(6):1211-21.
- Foster PJ, Buhrmann RR, Quigley HA, Johnson GJ. The definition and classification of glaucoma in prevalence surveys. *Br J Ophthalmol* 2002;**86**:238-42.
- Vithana EN, Khor CC, Qiao C, et al. Genome-wide association analyses identify three new susceptibility loci for primary angle closure glaucoma. *Nat Genet* 2012;**44**:1142-6.
- Foster PJ, Oen FT, Machin D, et al. The prevalence of glaucoma in Chinese residents of Singapore: a cross-sectional population survey of the Tanjong Pagar district. *Arch Ophthalmol* 2000;**118**:1105-11.
- Merula RV, Cronemberger S, Calixto N. [Incidence of primary angle-closure glaucoma in the Glaucoma Service of the Sao Geraldo Hospital]. *Arq Bras Oftalmol* 2008;**71**:389-93.
- Qu W, Li Y, Song W, et al. Prevalence and risk factors for angle-closure disease in a rural Northeast China population: a population-based survey in Bin County, Harbin. *Acta Ophthalmol* 2001;**89**:e515-20.
- Amerasinghe N, Zhang J, Thalamuthu A, et al. The heritability and sibling risk of angle closure in Asians. *Ophthalmology* 2011;**118**:480-5.
- Kong X, Chen Y, Chen X, Sun X. Influence of family history as a risk factor on primary angle closure and primary open angle glaucoma in a Chinese population. *Ophthalmic Epidemiol* 2011;**18**:226-32.
- Kavitha S, Zebardast N, Palaniswamy K, et al. Family history is a strong risk factor for prevalent angle closure in a South Indian population. *Ophthalmology* 2014;**121**:2091-7.
- Lowe RF. Primary angle-closure glaucoma. Family histories and anterior chamber depths. *Br J Ophthalmol* 1964;**48**:191-5.
- Lowe RF. Primary angle-closure glaucoma. Inheritance and environment. *Br J Ophthalmol* 1972;**56**:13-20.
- Sihota R, Ghate D, Mohan S, et al. Study of biometric parameters in family members of primary angle closure glaucoma patients. *Eye* 2008;**22**:521-7.
- Spaeth GL. Gonioscopy: uses old and new. The inheritance of occludable angles. *Ophthalmology* 1978;**85**:222-32.
- Tomlinson A, Leighton DA. Ocular dimensions in the heredity of angle-closure glaucoma. *Br J Ophthalmol* 1973;**57**:475-86.
- Etter JR, Affel EL, Rhee DJ. High prevalence of plateau iris configuration in family members of patients with plateau iris syndrome. *J Glaucoma* 2006;**15**:394-8.
- Tu YS, Yin ZQ, Pen HM, Yuan CM. Genetic heritability of a shallow anterior chamber in Chinese families with primary angle closure glaucoma. *Ophthalmic Genet* 2008;**29**:171-6.
- Foster PJ, Baasanhu J, Alsirk PH, et al. Glaucoma in Mongolia. A population-based survey in Hovsgol province, northern Mongolia. *Arch Ophthalmol* 1996;**114**:1235-41.
- Rosengren B. Studies in depth of the anterior chamber of the eye in primary glaucoma. *AMA Arch Ophthalmol* 1950;**44**:523-38.
- Phillips CI, Storey JK. Glaucoma geometry. *Exp Eye Res* 1971;**11**:140-1.
- Törnquist R. Shallow anterior chamber in acute glaucoma; a clinical and genetic study. *Acta Ophthalmol Suppl* 1953;**39**:1-74.
- Talluto D, Feith M, Allee S. Simultaneous angle closure in twins. *J Glaucoma* 1998;**7**:68-9.
- Congdon N, Wang F, Tielsch JM. Issues in the epidemiology and population-based screening of primary angle-closure glaucoma. *Surv Ophthalmol* 1992;**36**(6):411-23.
- Wickremasinghe S, Foster PJ, Uranchimeg D, et al. Ocular biometry and refraction in Mongolian adults. *Invest Ophthalmol Vis Sci* 2004;**45**:776-83.
- Wong TY, Foster PJ, Johnson GJ, et al. The relationship between ocular dimensions and refraction with adult stature: the Tanjong Pagar Survey. *Invest Ophthalmol Vis Sci* 2001;**42**:1237-42.
- Aung T, Bowman R, Chew PT, et al. Genome-wide linkage scan for primary angle-closure glaucoma. *Invest Ophthalmol Vis Sci* 2003;**44**:3224.
- Sundin OH, Dharmaraj S, Bhutto IA, et al. Developmental basis of nanophthalmos: MFRP is required for both prenatal ocular growth and postnatal emmetropization. *Ophthalmic Genet* 2008;**29**:1-9.
- Ayala-Ramirez R, Graue-Wiechers F, Robredo V, et al. A new autosomal recessive syndrome consisting of posterior microphthalmos, retinitis pigmentosa, foveoschisis, and optic disc drusen is caused by a MFRP gene mutation. *Mol Vis* 2006;**12**:1483-9.
- Nair KS, Hmani-Aifa M, Ali Z, et al. Alteration of the serine protease PRSS56 causes angle-closure glaucoma in mice and posterior microphthalmia in humans and mice. *Nat Genet* 2011;**43**:579-84.
- Aung T, Lim MC, Wong TT, et al. Molecular analysis of CHX10 and MFRP in Chinese subjects with primary angle closure glaucoma and short axial length eyes. *Mol Vis*, 2008;**14**:1313-8.
- Aung T, Yong VH, Chew PT, et al. Molecular analysis of the myocilin gene in Chinese subjects with chronic primary-angle closure glaucoma. *Invest Ophthalmol Vis Sci* 2005;**46**:1303-6.
- Day AC, Luben R, Khawaja AP, et al. Genotype-phenotype analysis of SNPs associated with primary angle closure glaucoma (rs1015213, rs3753841 and rs11024102) and ocular biometry in the EPIC-Norfolk Eye Study. *Br J Ophthalmol* 2013;**97**(6):704-7.
- Yang J, Siqueira MF, Behl Y, et al. The transcription factor ST18 regulates proapoptotic and proinflammatory gene expression in fibroblasts. *FASEB J* 2008;**22**:3956-67.
- Jandrig B, Seitz S, Hinzmann B, et al. ST18 is a breast cancer tumor suppressor gene at human chromosome 8q11.2. *Oncogene* 2004;**23**:9295-302.
- Nongpiur ME, Khor CC, Jia H, et al. ABCC5, a gene that influences the anterior chamber depth, is associated with primary angle closure glaucoma. *PLoS Genet* 2014;**10**:e1004089.

- Khor CC, Do T, Jia H, et al. Genome-wide association study identifies five new susceptibility loci for primary angle closure glaucoma. *Nat Genet* 2016;**48**:556-62.
- Othman MI, Sullivan SA, Skuta GL, et al. Autosomal dominant nanophthalmos (NNO1) with high hyperopia and angle-closure glaucoma maps to chromosome 11. *Am J Hum Genet* 1998;**63**:1411-8.
- Arner ES, Holmgren A. Physiological functions of thioredoxin and thioredoxin reductase. *Eur J Biochem* 2000;**267**:6102-9.
- Huber JC, Schneeberger C, Tempfer CB. Genetic modeling of estrogen metabolism as a risk factor of hormone-dependent disorders. *Maturitas* 2002;**41**(Suppl 1):S55-64.

### Further reading

Contact your local Allergan and ATHENA (Allergan Ophthalmic Professionals Educational Alliance) representative for the Early Diagnosis Program 4 – Gonioscopy in the diagnosis of glaucoma by Sancy Low and Gus Gazzard to get teaching materials and arrange departmental gonioscopy learning.

### TAKE HOME MESSAGE

- Genetic aetiologies are different for PACG and POAG.
- Older female first degree relatives are at highest risk of developing disease, with small ocular biometry and physical size as key contributing factors.
- Glaucoma and gonioscopic examinations can be recommended to patients who have AAC or PACG.
- Hyperopic refractive error overlaps with PACG, and may have shared genetic mechanisms.
- When a patient with atypical ACG presents to your clinic, consider connective tissue disorders.



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